Hall's Modified Harris Hematoxylin

Dolores W. Carter
Baptist Memorial Hospital
Memphis, Tennessee 38146

This modification of Harris' hematoxylin was discovered quite by accident by my supervisor, Sue Hall, who found that the presently available hematoxylin crystals dissolve more readily in water than in alcohol. This finding has made it possible for personnel in this laboratory to compound hematoxylin solutions (ready for use) in a few minutes. It has also eliminated the problem, which we have encountered and suspect others have also, of dissolving hematoxylin in alcohol. Our feeling is that this problem surfaced subsequent to the hematoxylin shortage of a few years ago, although we have no specific information to substantiate this opinion.

A search of the literature revealed that most Harris hematoxylin solutions require dissolving of the hematoxylin in absolute alcohol. In no instance did we find a method calling for water as the solvent. We feel, therefore, that this is a new way of compounding Harris' hematoxylin. It is presented here in the hope that other technicains may find it as useful and trouble-free as we have.

It must be noted that the hematoxylin and eosin stained slides using this hematoxylin have been excellent.

Hall's Modification of Harris' Hematoxylin

<table>
<thead>
<tr>
<th>Component</th>
<th>Quantity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hematoxylin crystals</td>
<td>5.0 gm</td>
</tr>
<tr>
<td>Ammonium or potassium alum</td>
<td>100.0 gm</td>
</tr>
<tr>
<td>Distilled water</td>
<td>1000.00 ml</td>
</tr>
<tr>
<td>Mercuric oxide (red)</td>
<td>2.5 gm</td>
</tr>
</tbody>
</table>

Dissolve the alum in the distilled water by the aid of heat. Add the hematoxylin crystals. Bring to a boil as rapidly as possible. Limit the boiling to less than one minute. Stir often during the boiling process. Remove from heat and add the mercuric oxide slowly. Reheat to a simmer until it becomes dark purple. Remove from heat immediately and place directly into a basin of cold water until solution is cool. The stain is ready for use as soon as it cools. Addition of 2-4 mL of glacial acetic acid per 100 mL of solution increases the precision of the stain. Filter before use.

The hematoxylin solution cited above is used in the conventional manner.

CAAMA Regional Program

Advances in Histopathology will be presented May 29-31, 1980, at the Beth Israel Medical Center in New York City, by the American Society of Clinical Pathologists.

Plastic is the "Light" Way to Go will be presented on June 13, 1980, at the Educational Center of the American Society of Clinical Pathologists in Chicago, Illinois.

For further information, contact: CAAMA Regional Program Manager; ASCP; 2100 W. Harrison St.; Chicago, IL 60612; (312) 738-1336, Ext. 154.

?? — Am I Intently Involved

An Editorial

The word INTENT is defined by Webster as: (1) Firmly fixed; earnest; intense. (2) Having the mind or attention firmly directed or fixed; engrossed, as he was intent on his studies.

This is a question which should be upper-most in our minds as Histotechnologists. The illustration shows a man walking on the sidewalk. He does not have to be intently involved on where he places every step. On the other hand, the man walking a one-foot plank over a deep ravine must be intently involved on where he places every step.

We as Histotechnologists must be intently involved in all facets of our daily involvement in the histopathology laboratory. Successful, high-quality slide production requires your full, continuous attention!

Illustration drawn by SSgt. Mitchel Duran, USAF.
Processing Aqueous Taps and Vitrectomies

Virginia Havener
Ophthalmic Pathology Laboratory
University of Minnesota
Minneapolis, Minnesota 55455

Recent advances in ophthalmic surgery have included the vitrectomy procedure, and, much earlier, the aqueous tap. Because of the minute amount of material obtained from these procedures, other methods were necessary to produce adequate slides for diagnosis. In our laboratory, various methods were investigated. They included centrifuging the specimen with subsequent filtration, paraffin embedding and sectioning. This process produced many artifacts (i.e., fibers of filter paper becoming enmeshed in the specimen, loss of some elements due to the chemicals and heat, etc.). Millipore filters were used but the process was time-consuming and the filters obscured cellular detail as well as posing problems for photography.

The Shandon Cytospin SCA-0030* solved most of the problems arising from scanty specimens and good artifact-clear slides can be obtained from as little as 0.5 ml of material using this unique instrument. Furthermore, the specimen can be processed fresh or fixed, and the method also allows procedures for determining the presence of fats, enzymes, or other substances which are usually lost by paraffin methods.

The Cytospin produces a monolayer of well-separated cells since they spin out according to weight of the cells. There is almost no distortion, so the cellular detail is excellent and the staining is vivid, resulting in a slide which is a pleasure to scan.

Following is the procedure presently used in this laboratory for processing vitrectomy and aqueous tap specimens:

1. If more than 10 ml of fluid are present with the specimen, spin down in a regular centrifuge and combine the resulting residues into a 10-ml sample or less.
2. If the amount of sample submitted is extremely small and four or more slides are desired, dilute the sample with up to 10 ml of 10% neutral-buffered formalin.
3. Fill the Cytospin head with clean blotters and slides which are alcohol cleaned.
4. Using a 1-ml pipette graduated in 0.1 ml, insert a maximum of 0.7 ml of specimen fluid into each well. (In order to get representative sample, be sure to shake bottle before each insertion.)
5. Place cover on head.
6. Set speed at 1200 rpm.
7. Turn right hand dial to 10. The green light will go on, to indicate cover is locked.
8. After a 10-minute spin, remove slides, etch accession number and place on warming plate until ready to stain — at least one hour. (If in a rush, a Gram and Giemsa stain may be performed after 10 minutes.) For the stains most frequently used at present, no further fixation is necessary, except for the Giemsa stain which after drying is pre-fixed with methanol.
9. For each vitrectomy specimen, we stain one slide with Gram and one with Giemsa, two H&E's, and other stains as desired or requested.
10. After using, clean the wells with Zephiran Chloride and allow to dry well before using again.

Note: Since it is necessary to have clean, dry wells, it is recommended that three sets of the wells be ordered. This will allow one set to be used while the others are drying.

This procedure is adaptable to any body fluids, including bone marrow specimens. The user's manual provides further information on the care and operation of the instrument.

*Shandon Southern Instruments, Inc.
Sewickley, PA 15143

Preliminary Program
ASMT Annual Meeting
June 22-27, 1980
St. Louis, Missouri

Histology

June 23
Working with Stained Glass
William Austin, DVM
Muscle Biopsy Procedures
Nathan Brinn
Duke University Medical Center

June 24
Steroid Receptors
Leslie Kane
University of Louisville Cancer Center
Legionnaires Disease
Patricia Greer
Center for Disease Control
Glycol Methacrylate
Nathan Brinn
Duke University Medical Center

June 25
Gross Dissection of the Rat as an Anatomical, Histologic and Histochemical Model
John Koski
McNeil Laboratories
Histopathology — Present and Future
J. Phillip Pickett
Duke University Medical Center
Cytology Preparatory Techniques
Gary Gill
John Hopkins University

June 26
Comparison of Computerized Tomography Scans with Gross Histologic Specimens
Richard Spencer & Alicia McKown
Jewish Hospital of Louisville
Advanced Optics
Hal Simpson
American Optical Corp.
Methyl Green Pyronin Staining Techniques
Sue Beth Landrum
University of Alabama Hospital
Immunohistochemical Approaches to the Lymphoid System
Richard Ford, M.D.
M. D. Anderson Hospital & Tumor Institute

An additional six lectures are scheduled during the week; however, titles were not available for this printing. For registration information, contact: ASMT; 330 Meadowfern Dr.; Houston, TX 77060.
National Society for Histotechnology Symposium
October 27-31, 1980
Atlanta, Georgia

The Sixth Annual Symposium/Convention of the National Society for Histotechnology will be conducted at the Colony Square Hotel, Atlanta, Georgia. The enclosed program is complete with hotel reservation card and registration form. The convention will utilize all sleeping accommodations in the Colony Square, with overflow accommodations in the Riviera Hyatt House. The Riviera Hyatt House is 5 minutes from our headquarters hotel. Room charges are the same at both hotels. All room reservations will be processed through the Colony Square. When the Colony Square is filled, reservations will be forwarded to the Riviera Hyatt House. The Riviera Hyatt House will provide shuttle service each morning and evening between their hotel and the Colony Square. Please make your reservations early since all rooms blocked for NSH will be released one month prior to our meeting date.

Mail hotel reservation directly to: Colony Square, Peachtree and 14th Street, Atlanta, GA 30361.

Symposium registration application may be photocopied if more than one individual from the same activity wishes to attend. To avoid delays and unnecessary complications, registrations awaiting fund approval will be accepted during the final convention month. Please include a note to this effect on your registration form. Mail registration and check to: NSH, P.O. BOX 36, LANHAM, MD 20861.

Meeting Schedule and Evening Activities

<table>
<thead>
<tr>
<th>Activities</th>
<th>Date</th>
<th>Time</th>
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<tr>
<td>Board of Directors</td>
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<tr>
<td>Meeting</td>
<td>Sun., Oct. 26</td>
<td>9 AM - 5 PM</td>
</tr>
<tr>
<td>Workshops</td>
<td>Mon. &amp; Tues., Oct. 27 &amp; 28</td>
<td>8:30 AM - 4:30 PM</td>
</tr>
<tr>
<td>Exhibits Open</td>
<td>Tues., Oct. 26</td>
<td>7 - 9 PM</td>
</tr>
<tr>
<td>Open Seminar: Diverse Topics in Research</td>
<td>Wed., Oct. 29</td>
<td>9:30 AM - 4 PM</td>
</tr>
<tr>
<td>Histology</td>
<td>Thurs., Oct. 30</td>
<td>8:30 AM - 10:30 AM</td>
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<tr>
<td>Open Seminar: How to Plan and Publish a State Newsletter</td>
<td>Tues., Oct. 28</td>
<td>1 - 4:30 PM</td>
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<tr>
<td>Scientific Sessions</td>
<td>Wed., Thurs., Fri, Oct. 29, 30, 31</td>
<td>8 AM - 4:30 PM</td>
</tr>
<tr>
<td>Exhibitors Liaison</td>
<td>Wed., Oct. 29</td>
<td>1 PM</td>
</tr>
<tr>
<td>Committee Meeting</td>
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NSH Membership Meeting    Wed., Oct. 29 4:45 - 6 PM
Career Awareness          Wed., Oct. 29 8 - 10 PM
Presentation Workshop     Wed., Thurs., Fri, Oct. 29, 30, 31 7 - 9 AM
Thomas Edison Exams       Thurs., Oct. 30 6 - 7 PM
Lab-Tek Cocktail Hour     Thurs., Oct. 30 7 - 10 PM
NSH Banquet               Sat., Nov. 1 9 AM
House of Delegates Meeting |

NSH Thomas Edison Program Monday - October 27:
Review sessions will be conducted from 9:00 AM to 4:00 PM for the following:
INTRODUCTORY HISTOTECHNOLOGY/HISTOCHEMISTRY
(Richard Schroeder)
CURRENT CONCEPTS IN DIAGNOSTIC HISTOPATHOLOGY
(Jules Elias)
9 AM - 12 NOON: HUMAN MICROSCOPIC ANATOMY
(Tom Palmer, Ph.D.)

Tuesday - October 28:
Review session from 9 AM to 4 PM for:
HUMAN MICROSCOPIC ANATOMY
(Freida Carson, Ph.D.)
7 - 10 PM CHEMISTRY: If registrant has pre-paid and will definitely be taking the Chemistry examination, this review session will be given. However, if no one will be taking the exam, review session will not be presented. You must prepay for this examination before coming to the symposium/convention in Atlanta.

There is no charge for attending review sessions.

COURSE EXAMINATIONS WILL BE ADMINISTERED THREE MORNINGS TO ALLOW PARTICIPANTS TO TAKE MORE THAN ONE EXAM DURING THE WEEK. EXAMS ARE SCHEDULED WEDNESDAY, THURSDAY AND FRIDAY, 7:00 - 9:00 A.M. Highlands Room.

COLONY SQUARE HOTEL
Attention: Reservations
Peachtree & 14th Streets, NE
Atlanta, Georgia 30361

NATIONAL SOCIETY FOR HISTOTECHNOLOGY
October 25 - November 2, 1980

I will arrive on ___________________ (Day) ___________________ (Date) and depart on ___________________ (Day) ___________________ (Date)

Name: _______________________

If sharing room, name of other occupant: _______________________

Mailing Address: _______________________

City: _______________________
State: _______________________
Zip: _______________________

To hold room after 6 p.m., indicate method of guarantee: □ One nights deposit enclosed $_____
(please circle type)

□ Credit Card: BAC/V - AE - MC - DC - CB Acct. No. __________ Exp. Date ______

Signature (required): _______________________

ACCOMMODATIONS REQUESTED:

Single: $40.00
Twin/Double: $48.00
Triple/Quad: $55.00
Reservations to be received no later than September 26, 1980

Reservations are cancelled after 6 p.m. on day of arrival unless secured by at least one night's deposit per room or accompanied by a written guarantee of payment for the first night's stay. No refund of the deposit will be made if cancellation is made less than 24 hours in advance of arrival date. This reservation request must be received 30 days in advance of arrival date. Room rates are subject to applicable taxes.
Check or money order must accompany registration fee for NSH Symposium. No cash or CODs will be accepted. Registration fee is due October 15th. No refunds for unattended workshops or seminars will be made after this date. Participants must be accompanied by a letter of registration. Fee includes meeting registration and membership in the Society. Registration fee for NSH Symposium/Congress members will be $80. No checks will be accepted for NSH Symposium/Congress members.

Non-NHS members must add $5.00 for each workshop and $10.00 for the scientific sessions. Canadian registrants please remit fees in U.S. currency.

Please Check: Is this your first attendance to an NSH Symposium/Congress? Yes No

Are you an NSH Member? Yes No

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Workshops

Monday, October 27, 1980

No. 1: Immunofluorescence
C.F. Craddock
8:30 AM - 9:30 AM
This workshop will cover the current theory and practice of immunofluorescent techniques in the routine and experimental histopathology laboratory. We shall briefly review the current theories of immunity, methods available for frozen and paraffin embedded tissues, and briefly discuss the use of the peroxidase-antiperoxidase (PAP) technique as a supplementary or alternate technique.

No. 2: Self-Assessment of Special Staining Techniques
(Donna Sheehan)
9:00 AM - 10:00 AM
This self-assessment workshop will give participants the ability to recognize various special staining techniques. They will understand the mode of action of various special stains with color results using photomicrographs. Demonstration of good quality control will be provided, including those tissues that are natural controls for various special stains. Discussion of the mode of action should make the participant aware of the sources of error and how they may be avoided. A Pre and Post test will be available to the participants.

No. 3: Micrometry Knife Sharpening
(George Harrison & James Harrison)
9:30 AM - 10:30 AM
Primary objective of this workshop is to instruct the participants in good knife sharpening. A number of knife sharpeners will be demonstrated along with a slide presentation. Bring your problem knives!

No. 4: Histology for Histotechnologists
(Margaret E. Wall, M.D.)
9:30 AM - 10:30 AM
Representative normal tissues have been cut from most body sites. Serial sections have been cut for routine and special stains. The histotechnologist will be led from an H&E in the appearance of the same area in each of the special stains. The goal is better documentation of well stained control slides.

No. 5: Laboratory Mathematics for Histotechnologists
(Jack M. Wang)
10:30 AM - 11:30 AM
This workshop will cover the mathematics dealing with normal, roller buffers and related solutions. The discussion will include conversion factor; i.e., preparation of dilute solution from one which is stronger, mathematical rules and examples, pH and pH factors pertinent to lhonenheit degree conversion. Numerous other items necessary in the preparation of solutions in the Histopathology Laboratory will be discussed. Weak and strong electrolytes will also be discussed if time permits.

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WORSHOPS

Monday

No. 1 $40 (all day)
No. 2 $40 (all day)
No. 3 $40 (all day)
No. 4 $40 (all day)
No. 5 $20 (½ day AM)
No. 6 $20 (½ day AM)
No. 7 $20 (½ day AM)
No. 8 $20 (½ day PM)
No. 9 $20 (½ day PM)
No. 10 $20 (½ day PM)

Tuesday

No. 11 $40 (all day)
No. 12 $40 (all day)
No. 13 $40 (all day)
No. 14 $40 (all day)
No. 15 $20 (½ day AM)
No. 16 $20 (½ day AM)
No. 17 $20 (½ day AM)
No. 18 $20 (½ day PM)
No. 19 $20 (½ day PM)
No. 20 $20 (½ day PM)
No. 21 $20 (½ day PM)

No. 6: Staining Characteristics of Legionella Pneumophila
(Patricia Grover & Stan Van Orden)
9:30 AM - 12 Noon
Limit: 25
This workshop will introduce the participants to the specific staining characteristics of Legionella pneumophila, focusing on Gram staining techniques. Participants will learn the importance of proper technique and the implications of different staining results.

No. 7: Quality Sections from Paraffin Embedded Tissues
(Mary King)
9:30 AM - 12 Noon
Limit: 25
This lecture will cover the techniques and considerations for preparing high-quality sections from paraffin-embedded tissues. Participants will learn how to optimize sectioning procedures to achieve best results.

No. 8: Professional Burnout
(Beverly Lynker, M.Ed., B.A.)
1:00 PM - 4:00 PM
Limit: 25
This workshop will address the issue of burnout in the workplace. Participants will learn strategies for managing stress and maintaining a healthy work-life balance.

No. 9: Self-Assessment Cytology - GYN
(Elizabeth Platt & Freda Martin)
1:00 PM - 4:00 PM
Limit: 25
This workshop will focus on self-assessment techniques for gynecologic cytology. Participants will practice their skills in identifying and analyzing cytologic specimens.

No. 10: Electron Microscopy for Histotechnologists
(Freda Curreri, PA.D.)
1:00 PM - 4:00 PM
Limit: 25
This workshop will introduce participants to the principles of electron microscopy as applied to histotechnology. Participants will learn how to interpret electron micrographs and understand their role in diagnostic pathology.
Tuesday, October 28, 1980

No. 11: Tissue Identification
(Leo G. Laux & Edwin Proskocilik)
8:30 AM - 9:30 PM
Primary objective of this workshop is to give each participant a basic knowledge of the microscopic structures of several of the commonly processed organs in the histopathology laboratory. It is anticipated that each histotechnologist will be sufficiently motivated to do further study. A handout of each slide of histology tissue will be given. This handout can then be applied to determining properly stained slides. In addition to learning the histotechnologist will be taught how to recognize proper staining qualities of numerous special stains.

No. 12: How to Plan an Experiment, Write a Scientific Paper and Present Data at a Scientific Meeting (P.A. Culling)
8:30 AM - 9:30 PM
This will be a hands-on workshop where participants will actually write a paper for a publication. One participant will be selected and placed on the Scientific Session Program for Friday, to present their paper written during this workshop.

No. 13: Glycol Methacrylate and Other Water Soluble Embedding Media
(Frank McCall)
8:30 AM - 9:30 PM
Limit: 40
Workshop will involve processing, embedding, sectioning and staining of water soluble plastics. Sections of plastic on glass and steel knives will be discussed and demonstrated. Low temperature processing with ultraviolet polymerization for enzyme histochemistry will also be discussed.

No. 14: Specimen Photography
(Robert Kershaw, E. Lewis Car & Jerry Binsker, Trix Photograph)
8:30 AM - 9:30 PM
The photography of specimens and slides is not a simple process. Basic microscope, use, alignment, cleaning, materials, films, glass slides, specimen preparation, photomicrography, brightfield, phase contrast, color, fluorescence, and special effects. There will be an hour of demonstration slides and practical slides. The afternoon session will deal with how to set up and operate your own laboratory within your hospital. Techniques on developing film will be discussed. Pictures taken during workshop will be developed during this time.

No. 15: Impact of Good Laboratory Practice Regulations on Histology Laboratories
(Donald B. Davis & J. D. P. Johnson)
8:30 AM - 12 Noon
A review of the ASAI and EPA Good Laboratory Practice regulations. Discussion of specific issues and examples of compliance of histology laboratories with these regulations. Clinical research: Requirements for standard office procedures; requirements for documentation of equipment maintenance; effects of compliance and non-compliance; and retrieval of raw data; reporting requirements; the relationship of the histology laboratory with the quality assurance unit.

No. 16: Immunoassay in Diagnostic Pathology
(Bob Jones)
8:30 AM - 12 Noon
The introduction of immunohistochemical methods has greatly increased the efficiency of the diagnostic process. As a result, the evaluation of the immunohistochemical techniques used will become essential. This workshop will be devoted to the evaluation of the immunohistochemical techniques used for the diagnosis of a variety of diseases.

No. 17: Microwave Fixation: A Routine Method for Rapid Fixation in a Surgical Pathology Laboratory
(Edward J. F. E. A. Johnson)
8:30 AM - 12 Noon
Workshop will consist of a combination of lecture and demonstration for the practical use of microwave fixation in routine histology. This method of fixation has been used for nearly two years as an alternative to formalin fixation and has been used in place of and in conjunction with formalin fixation. The result has been complete replacement of formalin in many instances and a reduction in fixation time from overnight to under three hours. This workshop will be devoted to the evaluation of microwave fixation.

No. 18: Analytical Histochemistry
(Frank Johnson, M.D.)
8:30 AM - 12 Noon
Limit: 50
There will be a presentation of simple, effective procedures for the recognition of inorganic species in tissue sections. There will be special emphasis on microcristallization.

No. 19: The Histochemistry Supervisor and the Interview
(Leo G. Laux, M.A. HT/ASCP)
1:00 PM - 3:00 PM
Much of the process of evaluating a candidate for a histotechnologist position requires a person to be interviewed. Workshop discussions will include some basics of laboratory management and will concentrate heavily on interviewing techniques. As managers of histopathology laboratories, you will be faced with the task of selecting individuals for interviewing. This workshop will be devoted to the evaluation of interviewing techniques. The evaluation of interviewing techniques will be presented by the workshop leader, followed by a panel discussion. The panel discussion will be geared to applying interviewing techniques to the laboratory setting, using student participants and experiences.

No. 20: Cytostomy
(Frank A. Ameline, B.A.)
1:00 PM - 3:00 PM
This portion of the workshop is to instruct the histotechnologist, novice and experienced, and associated individuals in the fine art of cytostomy positive forceps. This includes a brief synopsis of frozen sectioning techniques and the instrumentation used.

The integral part of the workshop will deal with the art of cytostomy itself. Discussions on tissue processing, use of materials and equipment and associated problems. Also, how to shop for new equipment and basic maintenance of existing equipment.

Basic staining of sections using routine stains, H&E, fat stains, histochromatographic methods, acid and alkaline phosphatase and immunofluorescence techniques. Discussion of kidney, skin, muscle biopsies, proper handling and collection. Crystallins will be available for class use.

No. 21: Proceedings on the Second Basic Science Workshop in Histology
(Morton Willsen, M.A. & Jules Ellis, M.A.)
1:45 PM
The following papers will be presented in this workshop:

1. Controls in Immunohistochemistry - Methods, Sensitivity, and Specificity: Controls for immunohistochemistry may be classified into two categories: (a) specificity using antigen negative controls, and (b) specificity using antigen positive controls.

2. Cytologic Flourescent Phosphates: The roles of H&E and fluoresent controls in the accuracy and reliability of fluorescence microscopy.

3. Developmental Disturbances During Tooth Formation: The development of teeth is a complex process that involves many stages of tissue differentiation. In this workshop, students will be introduced to the basic principles of tooth development and be encouraged to apply these principles to clinical situations.

4. New Methods for Serum Markers for Demineralized and Mineralized Bone: This workshop will discuss new methods for the detection of serum markers for demineralized and mineralized bone. These methods are being developed to detect new markers of bone turnover and to monitor the effectiveness of bone turnover.

5. Enzyme Histochemistry for Diagnosing Leukemia: The use of specific enzyme histochemical stains as a method of identifying cells by their functional traits will be discussed. The subtyping of the tumor types of leukemia is classified as follows: A solid tumor, B solid tumor, C solid tumor, and D solid tumor. The workshop will focus on the use of histochemical stains to differentiate between these tumor types.
Scientific Sessions

Wednesday, October 29, 1980

A.M. Session:

Laboratory Studies of Legionnaire's Disease
Substitution of Lead Nitrate for Uranium Nitrate as Used in the Steiner Silver
How to Set up Your Own Photography Lab in a Small Hospital
Asbestosis: Vital Role of Histotechnology in its Identification
Muscle Biopsy Histochemistry and Special Staining

Martin Hicklin, M.D.
Claire Greene, HT (ASCP)
Mike Ayers, HT (ASCP)
Frederick Gilbert, Jr., M.D.
Susi Schwarz, HT (ASCP)

P.M. Session:

Developing an Approved School of Histotechnology
Hazards of Infection in the Histology Laboratory
Histopathologic Diagnosis of Fungus Diseases
Communication Between Cytotechs and Histotechs: Better Specimen Preps

Gerre Welles, HT (ASCP)
John Otis, M.D.
Francis Chandler, DVM, Ph.D.
Ann Clark, B.S. &
Fonda Martin, B.S.
Joyce Eaton
Walter Scott, Ph.D.

Quality Control Guidelines for the Histopathology Laboratory
An Objective Tool for Grading Student Histosides

Thursday, October 30, 1980

A.M. Session:

Preparing for Laboratory Surveys
The Utilization of a New Emerging Health Professional — The Pathologist
Assistants
Does the Oncologist Really Need to See Those Slides?
Histochemical Demonstration of Hepatitis B Antigen: Technical and Diagnostic
Considerations
Medical-Legal Aspects of Histopathology
The Relevancy of Histopathology and Clinical Methodology in USDA Field
Service Laboratories Serving Federal Meat and Poultry Inspection Programs

Billie Swisher, HT (ASCP)
Denis Akim, P.A.
Melvin Moore, M.D.
Barbara Tersolo, HT (ASCP)
John Feegel, M.D.

Karl Langheinrich,
DVM, M.S., B.S.

P.M. Session:

Update on Rabies in the U.S.
Cytogenetics
Forensic Pathology
Time Utilization

William Winkler, DVM
Jack Reidy, Ph.D.
Larry Howard, Ph.D.
Betty Devon

Friday, October 31, 1980

Meeting Equipment Needs Under Cost Containment
Immunohistochemistry
Dermatopathology — A Challenge for Excellence
The Frozen Muscle Biopsy — Technique and Interpretation

Paper to be Presented Which was Written During Workshop on Tuesday
Panel Discussion: The Impact of Federal and State Regulations on
Histopathology and Cytopathology

Ewing Barnett
John Langloss, DVM, Ph.D.
Harold Meltzer, M.D.
Barbara Herr, B.A.

Patricia Greer, B.S.,
Fonda Martin, B.S.,
Ann Clark, M. Ed,
Marilyn Gamble, HT (ASCP)
A Guide for Educational Resources in Histotechnology

The next three issues of Histology will contain the remaining portions of an extensive list of various educational aids applicable to the field of Histotechnology. The July issue will contain a list of "Journals and Publications"; the October issue will feature "Visual and Audio Aids"; and the January issue will provide a list of miscellaneous training aids. This portion will be titled "Other Gems for Histotechnology."

The suggestion to incorporate this information in Histology was made by Ms. Irma B. Mednicoff, New England Medical Center, Boston, Massachusetts. The enormous task of compiling most of this information was performed by Ms. Gerre G. Welles, University of Tennessee, Center for Health Sciences, Memphis, Tennessee.

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<th>Author</th>
<th>Title</th>
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<th>Publisher Address</th>
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<tr>
<td>John D. Bancroft</td>
<td>HISTOCHEMICAL TECHNIQUES 2nd edition</td>
<td>Butterworths &amp; Co.</td>
<td>(1) Butterworths &amp; Company 161 Ash Street Reading, Massachusetts 01867</td>
</tr>
<tr>
<td>Gerrit Bevelander</td>
<td>OUTLINE OF HISTOLOGY (1971)</td>
<td>C. V. Mosby Co.</td>
<td>(3) C. V. Mosby Company 11630 Westline Industrial Drive St. Louis, Missouri 63141</td>
</tr>
<tr>
<td>Alexander Kennedy</td>
<td>BASIC TECHNIQUES IN DIAGNOSTIC HISTOPATHOLOGY</td>
<td>W. B. Saunders Co.</td>
<td>(13) W. B. Saunders Co. 500 Market St. San Francisco, California 94104</td>
</tr>
<tr>
<td>C. Roland Leesoon &amp; Thomas S. Leesoon</td>
<td>HISTOLOGY (1976)</td>
<td>W. B. Saunders Co.</td>
<td>(13) W. B. Saunders Co. 500 Market St. San Francisco, California 94104</td>
</tr>
<tr>
<td>Thomas J. McHale &amp; Paul T. Witzke</td>
<td>PERCENT, RATIO, PROPORTION</td>
<td>Williams &amp; Wilkins Co.</td>
<td>(16) Little, Brown &amp; Co. 34 Beacon St. Boston, Massachusetts 02156</td>
</tr>
<tr>
<td>Fredrick A. Putt</td>
<td>MANUAL OF HISTOPATHOLOGICAL STAINING METHODS (1972)</td>
<td>Year Book Medical Publishers, Inc.</td>
<td>(6) Year Book Medical Publishers, Inc. 35 E. Wacker Dr Chicago, Illinois 60601</td>
</tr>
</tbody>
</table>
Cell Blocks from Specimens of Body Fluids

Brenda Cuevas  
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Our laboratory had previously centrifuged specimens in glass conical tip tubes; however, even with centrifuge and fixation times of up to 1½ hours, we were often unable to obtain a compact button. I have observed that by substituting plastic conical tip centrifuge tubes (Becton-Dickinson 2087) for glass centrifuge tubes, one can obtain a more compact cell button with a corresponding shorter fixation time. The reason for this phenomenon is unknown and any possible explanation of why this occurs would be welcomed. Send information to the editor.

Helpful Hint for SEM Fixation

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Technicians who do electron microscopy procedures may be interested to learn that specimens can be fixed for Scanning Electron Microscopy with 3% glutaraldehyde made up in 0.1 M cacodylic buffer (pH 7.0). The specimen may remain in the fixative for up to three days without noticeable damage. After the fixation period, wash specimen with three changes of buffer and refrigerate until further processing can be carried out.

This technique has proven to be very beneficial for specimens containing bacteria and fungi.